

Can salivary composition and high flow rate explain the low caries rate in children with familial dysautonomia?

E. Mass, DMD N. Gadoth, MD D. Harell, PhD A. Wolff, DMD

Dr. Mass is senior lecturer, Department of Pediatric Dentistry, The Maurice and Gabriela Goldschleger School of Dental Medicine, Tel Aviv, Israel; Dr. Gadoth is professor, Department of Neurology, Meir General Hospital, Kfar Saba, and Sackler Faculty of Medicine, Tel Aviv University, Israel; Dr. Harell is director, Laboratory of Clinical Biochemistry, Rabin Medical Center, Beilinson Campus, Petah Tiqva, Israel; Dr. Wolff is head, Department of Hospital Dentistry, Assuta Hospital and Maccabi Healthcare Services, Tel Aviv, Israel. Correspond with Dr. Mass at elimas@post.tau.ac.il

Abstract

Purpose: Extremely low caries rate and increased major salivary gland flow rate have been previously reported in children with familial dysautonomia (FD). The purpose of this study was to explore the possibility that, in addition to increased salivary flow, children with FD have alterations in their salivary components, which may suggest an explanation to their low caries rate.

Methods: Whole unstimulated and stimulated saliva samples were collected from 13 children with FD who were found to be caries free, and from 28 age- and ethnic-matched healthy children, 15 caries-free children and 13 caries-affected children. The electrolyte and protein content of the unstimulated saliva and the microbial count and buffering capacity of the stimulated saliva were determined.

Results: Children with FD had the highest salivary flow rate and the lowest levels of mutans streptococci and lactobacilli, as well as the lowest concentration of chloride, magnesium, total protein and IgA. Healthy caries-affected children displayed the highest mutans streptococci and lactobacilli levels and lysozyme concentration, concomitantly with the lowest potassium and calcium concentrations.

Conclusions: The results of this investigation suggest that the caries-free state in FD may be associated with high salivary flow rate, while in healthy children, low caries rate may be associated with high salivary calcium concentration.(*Pediatr Dent.* 2002;24:581-586)

KEYWORDS: SALIVA, CARIES, FAMILIAL DYSAUTONOMIA

Received April 10, 2002 Revision Accepted July 17, 2002

Panilial dysautonomia (FD) is an autosomal recessive disorder that exclusively affects children of Jewish Ashkenazi origin. Clinical features such as impaired pain perception, skeletal deformities, recurrent pneumonias, orthostatic hypotension, progressive degeneration of tongue fungiform papillae and taste buds, impaired taste and smell, and multiple dysautonomic crises are the result of hereditary autonomic and sensory neuropathy (HASN) type III.¹ In a previous study, we have shown that children with FD are rarely affected by dental caries, despite their poor oral hygiene habits.² In subsequent studies, it was shown that children with FD have an increased major salivary gland flow rate and submandibular/sublingual gland hyperfunction at the acinar and ductal level, possibly due to denervation supersensitivity.^{3,4} FD is the first disease to cause basal salivary gland hyperfunction. Although the association of low salivary flow rate with increased caries activity is well known,^{5,6} there is no published evidence for the influence of high salivary flow rate on caries activity. Among the many caries-related salivary parameters are buffering capacity, mutans streptococci (MS) and lactobacilli (LB) levels,^{5,7} and concentrations of IgA, lysozyme,⁸ calcium, and phosphorus.^{9,10} However, correlations between single salivary protective constituents and caries rate are difficult to find.⁸

The purpose of this study was to analyze various salivary parameters in a group of children with FD and in 2 ageand ethnic-matched control groups (ie, caries affected and caries free).

Methods

The study population consisted of 41 Ashkenazi Jewish children divided into 3 groups: (1) The study group consisted of 13 patients with FD. (2) The control caries-free (CCF) group consisted of 15 healthy subjects. (3) The control caries-affected (CCA) group consisted of 13 healthy subjects. All 3 groups were pair matched by age and gender and came from a similar socioeconomic background. Patients with FD, who were randomly recruited from the FD patient database of 2 of the authors (EM and NG), fulfilled the diagnostic criteria for the disorder,¹¹ and the diagnosis was confirmed in each by one of the authors (NG). A detailed dental history was obtained with emphasis on oral hygiene habits and dietary characteristics, especially sugar intake and snack frequency. In addition, oral hygiene was assessed by the amount of bacterial plaque present on the teeth. Each of these parameters was qualified as "good," "moderate," or "poor." Specifically, up to 1 snack per day was considered a "good" dietary habit, 2 to 3 "moderate," and 3 or more "poor." Past exposure to fluoride was qualified as "significant" (persistent use of fluoridated drinking water and/or fluoride supplements) or "nonsignificant." After the receipt of informed consent by the parents and no more than 6 months prior to the saliva collection, patients and controls underwent a comprehensive oral examination by one of the authors (EM) who is an experienced pediatric dentist. The examination was a routine procedure in the examiner's private office and included bitewing radiographs.

Saliva collections were carried out between 9 AM to 12 PM. Subjects were instructed to refrain from eating, drinking, and tooth brushing for a minimum of 90 minutes before saliva collection. Unstimulated whole saliva was collected by expectoration into an ice-chilled tube for 5 minutes and its flow rate was calculated using gravimetric methods (assuming a specific gravity of saliva of 1.0) and expressed as ml/min.¹² For electrolyte and protein content measurements, these samples were frozen at -20°C and thawed shortly before analyses.

Stimulated whole saliva was collected following 5 minutes of paraffin chewing and immediately used for caries activity tests, for which commercially available methods (Orion Diagnostica, Espoo, Finland) were used. As children with FD were unable to chew effectively, reliable flow rates could not be obtained. Salivary buffering effect was assessed using the Dentobuff strip method, which defines a final pH of saliva of <4.5 as low (0), a pH of 4.5 to 5.5 as intermediate (1), and a pH >5.5 as high buffering activity (2).^{13,14} Salivary MS were tested with Dentocult SM Strip mutans test and their level was determined as scores 0, 1, 2, and 3 corresponding to <10,4 104-10,5 105-10,6 and >106 colonyforming units (CFU), respectively.¹⁵ Salivary LB were analyzed with Dentocult-LB dip-slide test, which classifies counts of $<10^{3}, 10^{3}, 10^{4}, 10^{5}, and \ge 10^{6}$ CFU as scores 0, 1, 2, 3, and 4.16,17

IgA and lysozyme were assayed by radial immunodiffusion on Kallestad Endoplate and Quantiplate kits,

Table 1. Mean Values (SD) of Salivary Parameters					
Parameter	FD*	CCF†	CCA‡		
Unstimulated flow rate (ml/min)	0.90 (0.66)	0.47 (0.27)	0.56 (0.22)		
Buffering capacity (U)	0.4 (0.7)	0.7 (1.3)	0.9 (1.0)		
Mutans streptococci (U)	0.2 (0.6)	1.5 (1.1)	2.2 (1.0)		
Lactobacilli (U)	0.5 (0.9)	1.3 (1.5)	3.1 (1.4)		
Sodium (mEq/l)	9.8 (3.2)	13.0 (5.2)	11.3 (3.0)		
Chloride (mEq/l)	11.0 (3.8)	18.0 (5.9)	13.2 (3.6)		
Potassium (mEq/l)	20.3 (5.7)	21.4 (4.1)	16.0 (3.4)		
Calcium (mg/dl)	2.2 (1.7)	2.5 (1.4)	1.5 (0.6)		
Phosphorus (mg/dl)	13.3 (7.2)	13.6 (3.7)	11.0 (3.4)		
Magnesium (mg/dl)	0.3 (0.1)	0.7 (0.5)	0.5 (0.3)		
Total protein (g/l)	1.1 (0.6)	1.7 (0.6)	1.9 (2.4)		
IgA (mg/dl)	2.4 (1.5)	3.4 (0.3)	3.2 (0.2)		
Lysozyme (mg/ml)	5.5 (6.4)	4.1 (1.8)	9.8 (8.0)		
Amylase (KU/l)	77.9 (54.4)	89.0 (93.2)	62.1 (49.4)		

*FD=familial dysautonomia

†CCF=control caries free

‡CCA=control caries active

respectively (Kallestad Laboratories Inc, Austin, Tex). Calcium and phosphorus were determined with a Hitachi 717 automated analyzer using Boehringer Mannheim kits. Magnesium was measured with a Perkin-Elmer model 2380 Atomic Absorption Spectrophotometer. Amylase was measured by the blocked PNG7 method using Raichem kit (RAI, San Diego, Calif). Sodium and potassium were measured with the IL 743 Flame Photometer (Instrumentation Laboratories Company, Lexington, Mass). Chloride was measured on a CMT 10 Chloridometer (Radiometer, Copenhagen, Denmark). Protein was determined according to Lowry.¹⁸

Statistical analyses were performed using one-way ANOVA procedures. Since multiple comparisons were carried out, the improved Bonferroni procedure was adopted.¹⁹

Results

The FD group was comprised of 10 boys and 3 girls aged 5 to 17 years (mean=10.5 years). The CCF group consisted of 11 boys and 4 girls (mean age=10.6 years), and the CCA group consisted of 10 boys and 3 girls (mean age 9.9 years). There was no statistically significant difference in the mean age of the groups. All were similar in terms of past exposure to fluoride, which was mostly "nonsignificant." The CCF and CCA children were similar in oral hygiene practices and dietary habits: 50% scored "moderate," 25% "good," and 25% "poor." Approximately 60% of children with FD had "poor" oral hygiene and altered dietary habits, 30% "moderate" and 10% "good." Most patients in the FD group were caries free, except 1 girl who had small carious

lesions on 2 proximal surfaces of adjacent primary molars and 1 boy who had 4 amalgam restorations on occlusal surfaces of primary and permanent molars. Therefore, the dental caries status of the FD group was considered similar to the CCF group. The mean overall DMFdef rate in the CCA group was 9.7±3.3.

The values of the measured salivary parameters are shown in Table 1 and the results of the statistical analysis in Table 2. Children with FD differed significantly from all controls combined together in terms of higher salivary flow rate and lower MS and LB counts, as well as decreased chloride, mag-

Groups compared	FD vs CCA	FD vs CCF	FD+CCF vs CCA	CCA vs CCF	FD vs CCA+CCI
Unstimulated flow rate	NS	<i>P</i> <.02	NS	NS	<i>P</i> <.01
Mutans streptococci	<i>P</i> <.0001	<i>P</i> <.0005	<i>P</i> <.002	NS	<i>P</i> <.0001
Lactobacilli	<i>P</i> <.0001	NS	<i>P</i> <.0001	<i>P</i> <.001	<i>P</i> <.005
Lysozyme	<i>P</i> <.005	NS	<i>P</i> <.001	<i>P</i> <.005	NS
Chloride	NS	<i>P</i> <.001	NS	NS	<i>P</i> <.01
Potassium	NS	NS	<i>P</i> <.005	<i>P</i> <.005	NS
Calcium	NS	NS	<i>P</i> <.02	<i>P</i> <.02	NS
IgA	NS	<i>P</i> <.01	NS	NS	<i>P</i> <.005
Magnesium	<i>P</i> <.0005	NS	NS	NS	<i>P</i> <.001
Total protein	NS	NS	NS	NS	<i>P</i> <.05

NS=not significant

No statistical significance was found between the study groups in the values of buffering capacity, sodium, phosphorus, and amylase

nesium, total protein, and IgA concentrations. Although both FD and CCF groups had a similar dental caries status (FD had an extremely low caries rate), salivary flow rates were higher in FD, while MS, chloride, and IgA concentrations were significantly lower. Saliva from the CCA group differed from that of FD and CCF groups regarding lower potassium and calcium concentrations and higher MS, LB, and lysozyme levels. Potassium and calcium were lower in the CCA group compared to the CCF group while MS and magnesium were higher in the CCA group, compared to the FD group. The CCA group had a consistently higher LB level and lysozyme concentration than FD and CCF groups. Output per minute of all analyzed salivary constituents did not differ significantly between the 3 groups.

Discussion

The primary goal of the present study was to characterize a salivary profile that could be responsible for the extremely low caries level found in children with FD. The most striking finding was the significantly increased salivary flow rate in FD compared to the combined CCA and CCF groups. As previously reported, salivary gland hyperfunction in FD, as manifested by the increased flow rate, may be due to denervation supersensitivity of the partially denervated salivary glands.³ Increased salivary flow rate may significantly contribute to oral health by optimizing cleansing and increasing buffering power and degree of saturation with respect to tooth minerals.^{6,8,20} Increased salivary flow rate was the only "caries protective" parameter found in FD as compared to the combined CCA and CCF groups. Although no statistically significant difference in salivary flow rate was found between the FD and the CCF groups, there was a trend towards higher flow rate in FD. Therefore, it may be assumed that, despite poor oral hygiene and altered dietary habits,² children with FD are protected from dental caries by their high salivary flow rate.

The CCF group had the highest salivary calcium and phosphorus concentrations. However, statistical significance was found only when the CCF group was compared with the CCA in respect to calcium concentration. The role of calcium and phosphorus in dental remineralization is well known,⁵ and their concentration in whole stimulated saliva reflects their level in plaque.²¹ High levels of calcium and phosphorus in whole and parotid saliva were found to be associated with a low dental caries rate.^{9,10,22} However, in other studies,^{23,24} no such association could be found. The crucial anticariogenic role of calcium was also evident in the present study, since the CCF children had a high salivary calcium concentration while other protective salivary parameters were similar to those of the CCA group.

The salivary MS concentration in FD was extremely low. There was also a tendency for lower MS in the CCF group compared with the CCA group. This is in line with previous studies showing a direct relationship between caries rate and MS levels.^{23,25-27} The low MS levels in FD saliva may be associated with the high salivary flow rate rather than with the antibacterial activity of IgA and lysozyme, which were not increased in the FD group. The association between salivary flow rate and MS levels is in agreement with previous studies on children and young adults.^{25,28}

The salivary lactobacilli level reflects the presence of bacteria harboring niches in the mouth, either iatrogenic or caused by carious cavities.²⁹ Since none of the children examined had niches secondary to orthodontic appliances or overhanging dental restorations, it was not surprising that the CCA group had the highest lactobacilli levels, due to carious cavities. Similarly to a previous report,³⁰ a dip-slide tested lactobacilli level proved to be of paramount importance in the discrimination between children with and without caries.

Salivary buffering capacity in FD was comparable to that reported for 11- to 12-year-old Finnish children with low (0.63-0.70) DMFS levels,²⁸ but was slightly lower than that reported in another Finnish study.³¹ Although buffering capacity was highest in CCA and lowest in FD, these differences did not reach statistical significance. This lack of significance may be due to the limited sensitivity of the methods used.

These simple, commercially available methods are widely used by many investigators to measure the buffering capacity and MS and lactobacilli levels in children.^{26-28,30-33} They have been recommended by a working group from the Commission on Oral Health, Research, and Epidemiology (CORE).⁵ The disadvantage of these methods is their semiquantitative nature, which limits the resolution power of statistical comparisons. Another contributing factor to the lack of significant differences in the results of buffering capacity was the difficulty in obtaining reliable amounts of stimulated whole saliva from children with FD, which may have interfered with the consistency of the values obtained.

Generally, the values for the various salivary components of the healthy control groups were similar to those previously reported in healthy chldren.^{10,12,21,22,31,32,34-40} In addition, as previously shown,³¹ the authors found a negative correlation between salivary flow rate and total protein content. Indeed, the FD group, while displaying the highest flow rate, had the lowest salivary total protein concentration. In contrast, the submandibular/sublingual protein concentration was found to be similar in FD and healthy control children.⁴

As stated in the Results section, several significant differences were found in regards to the concentrations of salivary components, but none concerning their output. The relevance of the concentrations' differences, as discussed in this section, will not be completely elucidated until confirmation or rejection of the present data regarding the output of salivary components will be obtained on a larger sample.

The role of IgA in oral immunologic defense against streptococcal infection has been considered part of the overall mucosal defense mechanism.^{8,40,41} Minor salivary glands, which are responsible for the secretion of approximately 30-55% of IgA, react to antigenic challenges in the oral cavity by producing IgA.^{40,42,43} The results of the present study may suggest that in FD, decreased IgA concentration reflects a reduced "need for protection," since the salivary infection level was low. An inverse relationship between caries experience and the concentration of IgA has been previously reported,⁴⁴ but the literature is inconsistent regarding the association between the caries rate and salivary IgA level.^{28,45} The finding of a low IgA concentration with a normal output is in line with the above mentioned inconsistency.

The CCA group had the highest value of salivary lysozyme, which belongs to the nonimmunologic salivary

defense system. This may suggest the presence of a feedback mechanism by which lysozyme is released into the oral cavity in response to the need of protection required against caries activity. However, in previous studies, no correlation was found between parotid and submandibular-sublingual whole salivary lysozyme concentrations and caries state.^{28,34,46}

No statistically significant differences were found between the 3 groups for amylase concentrations. Similarly, Dodds et al,⁴⁷ reported no differences in total protein and amylase concentrations in parotid saliva between caries-free and caries-active adults. Although amylase may interact with oral bacteria to provide glucose from dietary starch, little is known about the significance of this interaction with plaque formation or dental caries.^{48,49}

It is difficult to interpret the electrolyte values in the present study in view of lack of consensus as to their role in the caries process. While several investigators have reported higher potassium and chloride levels in parotid saliva from caries-active adults,^{47,50} others ruled out an association between these electrolytes and the caries rate.^{9,51} The authors' caries-active group had a significantly lower salivary concentration of potassium, while chloride was similar in both healthy control groups but lower among the FD children.

Salivary magnesium has been reported to be either unrelated^{9,23,24,51} or directly associated with caries rate.⁵⁰ In the present study, magnesium was similar in both healthy control groups and significantly higher among the FD children.

Conclusions

- 1. The results of the study surmise an anticariogenic role of increased salivary flow rate in FD, that needs to be confirmed by a larger study.
- 2. The high salivary concentration of calcium in CCF children seems to be a significant anticariogenic factor.
- 3. Salivary levels of mutans streptococci and lactobacilli were associated with the salivary flow rate and possibly with the degree of caries activity (since the CCA group displayed the highest values of these parameters).
- 4. Investigation of the role of other salivary factors in the dental caries status of healthy and FD children should be continued in further studies.

Acknowledgments

This study was supported by a grant from The Lefcoe Oral Biology Research Fund in Memory of Mavolyn-Brown Lefcoe. The authors wish to express their gratitude to Ms. Rita Lazar for her editorial assistance.

References

 Dyck PJ, Ohta M. Inherited neuronal degeneration and atrophy affecting peripheral motor, sensory and autonomic neurons; In: Dyck PJ, Thomas PK, Lambert EM, eds. *Peripheral Neuropathy.* Philadelphia, Pa: WB Saunders; 1975:791-824.

- Mass E, Sarnat H, Ram D, Gadoth N. Dental and oral findings in patients with familial dysautonomia. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 1992;74:305-311.
- 3. Mass E, Wolff A, Gadoth N. Increased major salivary gland secretion in familial dysautonomia. *Dev Med Child Neurol.* 1996;38:133-138.
- 4. Wolff A, Harell D, Gadoth N, Mass E. Submandibular/sublingual salivary gland function in familial dysautonomia. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2002;94:315-319.
- 5. Sreebny LM, Banoczy J, Baum BJ, Edgar WM, Epstein JB, Fox PC, et al. Saliva: its role in health and disease. *Int Dent J.* 1992;42(suppl 2):291-304.
- Edgar WM, Higham SM, Manning RH. Saliva stimulation and caries presentation. *Adv Dent Res.* 1994; 8:239-245.
- Brattall D, Carlsson J. Current status of caries activity tests. In: Thylstrup A, Fejerskov O, eds. *Textbook of Cariology*. 1st ed. Copenhagen, Denmark: Munksgaard; 1986:249-265.
- Tenovuo J. Antimicrobial function of human saliva– how important is it for oral health? *Acta Odontol Scand.* 1998;56:250-256.
- Shannon IL, Feller RP. Parotid saliva flow rate, calcium, phosphorus, and magnesium concentrations in relation to dental caries experience in children. *Pediatr Dent.* 1979;1:16-20.
- 10. Shaw L, Murray JJ, Burchell CK, Best JS. Calcium and phosphorus content of plaque and saliva in relation to dental caries. *Caries Res.* 1983;17:543-548.
- Gadoth N, Abramovitch D, Melamed E. The regional cerebral flow in familial dysautonomia. *Brain Dev.* 1989;11:179-182.
- Mason DK, Chisholm DM. Saliva. In: Mason DK, Chisholm DM, eds. *Salivary Glands in Health and Dis*ease. London, England: WB Saunders Company Ltd; 1975:37-69.
- 13. Frostell G. A colourimetric-screening test for evaluation of the buffer capacity of saliva. *Swed Dent J.* 1980;4:81-86.
- Ericson D, Bratthall D. Simplified method to estimate salivary buffer capacity. *Scand J Dent Res.* 1989;97:405-407.
- 15. Jensen B, Bratthall D. A new method for the estimation of mutans streptococci in human saliva. *J Dent Res.* 1989;68:468-471.
- Larmas M. A new dip-slide method for the counting of salivary lactobacilli. *Proc Finn Dent Soc.* 1975;71:31-35.
- Parvinen T, Larmas M. The relation of stimulated salivary flow rate and pH to lactobacillus and yeast concentrations in saliva. *J Dent Res.* 1981;60:1929-1935.
- Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ. Protein measurements with the Folin phenol reagent. *J Biol Chem.* 1951;193:265-275.

- 19. Simes RJ. An improved Bonferroni procedure for multiple tests of significance. *Biometrika*. 1986;73: 751-754.
- 20. Lagerlöf F, Oliveby A. Caries-protective factors in saliva. *Adv Dent Res.* 1994;8:229-238.
- 21. Ashley FP, Coward PY, Jalil RA, Wilson RF. Relationship between calcium and inorganic phosphorus concentrations of both resting and stimulated saliva and dental plaque in children and young adults. *Arch Oral Biol.* 1991;36:431-434.
- 22. Sewón SM, Mäkelä M. A study of the possible correlation of high salivary calcium levels with periodontal and dental conditions in young adults. *Arch Oral Biol.* 1990;35:211S-212S.
- Bowen WH, Velez H, Aguila M, Velasaues H, Sierra LI, Gillespie G. The microbiology and biochemistry of plaque, saliva and drinking water from 2 communities with contrasting levels of caries in Colombia, SA. *J Dent Res.* 1977;55(special issue C):C32-C39.
- 24. Borella P, Fantuzzi G, Aggazzotti G. Trace elements in saliva and dental caries in young adults. *Sci Total Environ*. 1994;153:219-224.
- Gråhn E, Tenuvuo J, Lehtonen O-P, Eerola E, Vilja P. Antimicrobial systems of human whole saliva in relation to dental caries, cariogenic bacteria, and gingival inflammation in young adults. *Acta Odontol Scand.* 1988;46:67-74.
- Granath L, Cleaton-Jones P, Fatti LP, Grossman ES. Prevalence of dental caries in 4- to 5-year old children partly explained by presence of salivary mutans streptococci. *J Clin Microbiol.* 1993;31:66-70.
- 27. Llena-Puy MC, Montañana-Llorens C, Forner-Navarro L. Cariogenic oral flora and its relation to dental caries. *ASDC J Dent Child*. 2000;67:42-46.
- Tukia-Kulmala H, Tenovuo J. Intra- and inter-individual variation in salivary flow rate, buffer effect, lactobacilli, and mutans streptococci among 11- to 12-year-old schoolchildren. *Acta Odontol Scand.* 1993;51:31-37.
- 29. Crossner C-G. Salivary lactobacillus counts in the predictions of caries activity. *Community Dent Oral Epidermiol.* 1981;9:182-190.
- Leverett DH, Proskin HM, Featherstone JDB, Adair SM, Eisenberg AD, Mundorff-Shrestha SA, et al. Caries risk assessment in a longitudinal discrimination study. *J Dent Res.* 1993;72:538-543.
- Söderling E, Pienihäkkinen K, Alanen M-L, Mietaoja M, Alanen P. Salivary flow rate, buffer effect, sodium, and amylase in adolescents: a longitudinal study. *Scand J Dent Res.* 1993;101:98-102.
- Pajari U, Poikonen K, Larmas M, Lanning M. Saliva immunoglobulins, lysozyme, pH, and microbial counts in children receiving anti-neoplastic therapy. *Scand J Dent Res.* 1989;97:171-177.
- 33. Gábris K, Nagy G, Madléna M, Denes Z, Marton S, Keszthelyi G. Associations between microbiological and

salivary caries activity tests and caries experience in Hungarian adolescents. *Caries Res.* 1999;33:191-195.

- MacKay BJ, Goodman H, Cox D, Grossbard BL, Iacono VJ, Pollock JJ. Development of an enzymelinked immunosorbent assay for determination of lysozyme in human parotid and submandibular-sublingual salivas. *J Clin Microbiol*. 1984;19:844-848.
- Grundbacher FJ. Variation in levels of immunoglobulins A, G, and E in human saliva. Arch Oral Biol. 1988;33:121-126.
- Olness K, Culbert T, Uden D: Self-regulation of salivary immunoglobulin A by children. *Pediatrics*. 1989;83:66-71.
- Tenovuo JO. Human Saliva: Clinical Chemistry and Microbiology. Boca Raton, Fla: CRC Press Inc; 1989: Vol. 1:9-17, 26-61, 75-94; Vol. 2:28-30, 104-108.
- Ben-Aryeh H, Fisher M, Szargel R, Laufer D. Composition of whole unstimulated saliva of healthy children: Changes with age. *Arch Oral Biol.* 1990; 35:929-931.
- 39. Jalil RA, Ashley FP, Wilson RF, Wagalyu EG. Concentrations of thiocyanate, hypothiocyanite, 'free' and 'total' lysozyme, lactoferrin and secretory IgA in resting and stimulated whole saliva of children aged 12-14 years and the relationship with plaque accumulation and gingivitis. J Periodont Res. 1993;28:130-136.
- Russell MW, Hajishengallis G, Childers NK, Michalek SM. Secretory immunity in defense against cariogenic mutans streptococci. *Caries Res.* 1999;33:4-15.
- McGhee JR, Michalek SM. Immunobiology of dental caries–microbial aspects and local immunity. *Annu Rev Microbiol.* 1981;35:595-638.

- 42. Crawford JM, Taubman MA, Smith DJ. Minor salivary glands as a major source of secretory immunoglobulin A in the human oral cavity. *Science*. 1975; 190:1206-1209.
- 43. Nair PNR, Schroeder ME. Duct-associated lymphoid tissues (DALT) of minor salivary glands and mucosal immunity. *Immunology*. 1986;57:171-180.
- 44. Lehner T, Cardwell JE, Clarry ED. Immunoglobulins in saliva and serum in dental caries. *Lancet*. 1967; 1:1294-1297.
- 45. Ørstavik D, Brandtzaef P. Secretion of parotid IgA in relation to gingival inflammation and dental caries experience in man. *Arch Oral Biol.* 1975;20:701-704.
- Stuchell RN, Mandel ID. A comparative study of salivary lysozyme in caries-resistant and caries-susceptible adults. *J Dent Res.* 1983;62:552-554.
- Dodds MWJ, Johnson DA, Mobley CC, Hattaway KM. Parotid saliva protein profiles in caries-free and caries-active adults. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 1997;83:244-251.
- Scannapieco FA, Torres G, Levine MJ. Salivary α-amylase: Role in dental plaque and caries formation. *Crit Rev Oral Biol Med.* 1993;4:301-307.
- 49. Dowd FJ. Saliva and dental caries. *Dent Clin North Am.* 1999;43:579-597.
- 50. Kargul B, Yarat A, Tanboga I, Emekli N. Salivary protein and some inorganic element levels in healthy children and their relationship to caries. *J Marmara Univ Dent Fac.* 1994;2:434-440.
- Shannon IL, Kilgore WI, Terry JM. Relation of parotid fluid flow rate, sodium, potassium, and chloride concentrations to caries experience. *J Oral Med.* 1969;24:3-5.

Abstract of the Scientific Literature

A CEPHALOMETRIC STUDY OF THE CLASS II CORRECTION EFFECTS OF THE EUREKA SPRING

The Eureka Spring appliance (ES) is a Class II treatment alternative for noncompliant patients and, in Herbst tradition, has a compressible plunger assembly attached posteriorly to maxillary molars and anteriorly to the mandibular cuspid/bicuspid region. ES is reported to be advantageous over other Class II treatment modalities in noncompliant patients. The purpose of this study was to evaluate cephalometric treatment effects in such cases. Cephalometric measurements were obtained for 37 growing and nongrowing Class II patients at 2 time intervals, including insertion and removal of ES and analyzed using paired *t* tests (P<.05). Class I and favorable overjet and overbite relationships were obtained in all patients in a mean treatment time of 4.0±1.3 months. Significant changes attributed to the ES were dental and included changes in maxillary and mandibular molar and incisor position, angulations, and/or inclination and orientation of the occlusal plane to Frankfort Horizontal attributed to intrusion of teeth. No significant skeletal changes were established. Thus, the authors conclude that there may be specific advantages to ES over standard Class II correction methods.

Comments: Despite the absence of either a control or comparison group, this study demonstrates possible dental changes using ES with limited skeletal effects. AW

Address correspondence to J. P. DeVincenzo, Department of Orthodontics, School of Dentistry, Loma Linda University, Loma Linda, Calif. jdev@digitalputty.com

Stromeyer EL, Caruso JM, DeVincenzo JP. A cephalometric study of the Class II correction effects of the Eureka Spring. *Angle Orthod.* 2002;72:203-210.